GenJet™ In Vitro DNA Transfection Reagent for MCF-7 Cells (Ver. II)

---- A Protocol for Transfecting MCF-7 Cells

100	μΙ
500	
1000	



15875 Gaither Drive Gaithersburg, MD 20877 FAX. 301-560-4919 TEL. 301-330-5966 Toll Free. 1-(866)-918-6812

Email: <u>info@signagen.com</u>
Web: <u>www.signagen.com</u>

This product is for laboratory research ONLY and not for diagnostic use

Introduction:

GenJet™ In Vitro DNA Tranfection Reagent (Ver. II) is upgraded version of GenJet™ In Vitro DNA Tranfection Reagent. With a new chemistry, more DNA condensing groups were released in the new version compared with old version GenJet™, leading to 3~4 times more efficient in DNA delivery. GenJet™ (Ver. II) for MCF-7 cells was Pre-optimized and conditioned for transfecting MCF-7 cells.

Procedures for Transfecting MCF-7 Cells: Step A. Cell Seeding (see <u>Table 1</u>):

Cells should be plated 18 to 24 hours prior to transfection so that the monolayer cell density reaches to the optimal $\sim 90\%$ confluency at the time of transfection. Complete culture medium with serum and antibiotics is freshly added to each well ~ 60 minutes before transfection.

Table 1. A Guideline for Seeding Adherent Cells Prior to Transfection in Different Culture Formats

Culture Dishes	Surface Area (cm²)	Number of Cells to Seed
T75 Flask	75	3.0 - 6.0 x 10 ⁶
100 mm Dish	58	2.2 - 4.4 x 10 ⁶
60 mm Dish	21	0.9 - 1.8 x 10 ⁶
35 mm Dish	9.6	3.5 - 7.0 x 10 ⁵
6-well Plate	9.6	4.0 - 8.0 x 10 ⁵
12-well Plate	3.5	1.5 - 3.0 x 10 ⁵
24-well Plate	1.9	0.8 - 1.6 x 10 ⁵
48-well Plate	1.0	4.0 - 8.0 x 10 ⁴
96-well Plate	0.3	1.2 - 2.4 x 10 ⁴

Table 2. Recommended Amounts for Different Culture Vessel Formats

Culture Dish	Transfection Volume (ml)	Plasmid DNA (μg)	Diluent Volume (mL)	GenJet™ Reagent (μL)
96-well	0.2	0.2	2 x 0.01	0.6
48-well	0.3	0.5	2 x 0.02	1
24-well	0.5	1.0	2 x 0.05	3
6-well	1.2	2	2 x 0.1	6
35 mm dish	1.5	2	2 x 0.1	6
60 mm dish	3	5	2 x 0.25	15
10 cm dish	6	7 - 8	2 x 0.5	21 - 24
T75 flask	10	18 - 36	2 x 0.75	54 - 108
250 ml flask	20	50 - 100	2 x 1.25	150 - 300

Step B. Preparation of GenJet™-DNA Complex and Transfection Procedures

For MCF-7 cells, the optimal ratio of GenJet™ (μL):DNA (μg) is 3:1. To ensure the optimal size of complex particles, we recommend using serum-free DMEM with High Glucose to dilute DNA and GenJet™ Reagent.

The following protocol is given for transfection in 24-well plates, refer to **Table 2** for transfection in other culture formats. The optimal transfection conditions For MCF-7 cells are given in the standard protocol described below.

- For each well, add 0.5 ml of complete medium with serum and antibiotics freshly $\sim\!60$ minutes before transfection.
- For each well, dilute 1 μg of DNA into 50 μl of serum-free DMEM with High Glucose. Vortex gently and spin down briefly to bring drops to bottom of the tube .
- For each well, dilute 3 µl of GenJet™ reagent (Ver.
 II) into 50 µl of serum-free DMEM with High Glucose.

 Vortex gently and spin down briefly.
- Add the diluted GenJet™ Reagent immediately to the diluted DNA solution all at once. (Important: do not mix the solutions in the reverse order!)
- Vortex-mix the solution immediately and spin down briefly to bring drops to bottom of the tube followed by incubation of 15 minutes at room temperature to allow GenJet™-DNA complexes to form.

Note: Never keep the DNA/GenJet[™] complex longer than 20 minutes

- Add the 100 µl GenJet[™]/ DNA complex drop-wise onto the medium in each well and homogenize the mixture by gently swirling the plate.
- Remove DNA/GenJet™ complex-containing medium and replace with fresh complete serum/antibiotics containing medium 12~18 hours post transfection.
- Check transfection efficiency 24 to 48 hours post transfection.

Storage: GenJet[™] DNA In Vitro Transfection Reagent is stable for up to 12 months at +4 °C. This item shipped at ambient temperature